

ViroVision™ HIV Reporter Cell System: Cell Culture Protocol

Important:

- 1) **Results are only guaranteed if ViroVision™ Infection Enhancement Medium is used when cells are infected with HIV.**
- 2) **Once thawed, the Infection Enhancement Medium should be stored at 4°C, and is stable for 3 months. Do not re-freeze and do not leave Infection Media at room temperature.**
- 3) **Depending on the cell line, there may be different media requirements. Please consult the table at the end to match the proper media with purchased cell line.**

Contents

Item	QTY	Storage
ViroVision™ Cells	~ 5 X 10 ⁶ Cells	Liquid N ₂
ViroVision™ Infection Enhancement Medium	200 µl of 10X solution	-20°C until needed. 4°C for up to 3 months once thawed.

Initiation of ViroVision™ Cell Culture

- 1) Please ensure cells arrive frozen. Contact Cube Biosystems immediately if cells arrive thawed.
- 2) Store cells in liquid N₂ until needed.
- 3) To initiate cell culture, remove a vial of frozen cells from storage, and thaw as quickly as possible by placing in a 37°C water bath.

Note: If cells are removed from liquid nitrogen, under sterile conditions, immediately unscrew cap ½ to 1 full turn to allow N₂ gas to escape. Re-secure cap. After cap is re-secured, place cells in 37°C water bath to thaw.

- 4) Pre-warm 5 ml appropriate complete media in 15 ml tube to 37°C. Add cells, dropwise, to pre-warmed media. Mix gently.
- 5) Collect the cells by centrifugation at 300 x g for 5 minutes, room temperature.
- 6) Remove/aspirate the supernatant and resuspend the cell pellet in 15 ml of complete media.
- 7) Place the cells into a T75 flask and incubate at 37°C, 5% CO₂.

ViroVision™ HIV Reporter Cell System: Cell Culture Protocol

Culturing ViroVision™ Cells

- 1) Count cells daily and keep at a density below 1×10^6 cells ml^{-1} . Dead cells may be removed by Ficoll separation.
- 2) Add fresh complete media when cell density reaches 1×10^6 ml^{-1} .

HIV Infection

- 1) Count cells and pellet cells by centrifugation at $300 \times g$ for 5 minutes.

Note: Cell viability should be $\geq 80\%$.

- 2) Resuspend cells in complete media at concentration of $\sim 2 \times 10^6$ cells ml^{-1} .
- 3) Use 100 μl of cells ($\sim 2 \times 10^5$) per infection.
- 4) Pre-treat cells by adding 10 μl of ViroVision™ Infection Enhancement Medium (10 x) so that Infection Media concentration is $\sim 1 \times$. Mix and incubate for 2 hours. Use of ViroVision™ Infection Enhancement Medium is required.
- 5) Add virus to the cells & mix. Note volume of virus used.
- 6) Add Infection Enhancement Medium to 1/10 of the virus volume used. E.g. If 100 μl of virus used, add 10 μl of Infection Enhancement Medium. Incubate at 37°C for 2-4 hours.
- 7) Add 1 ml fresh complete media to wash cells. Pellet cells as above and remove supernatant. (Optional) Repeat 1 x for a total of 2 washes.
- 8) After washing, resuspend cells in 1 ml complete media.
- 9) Culture cells at 37°C , 5% CO_2 for 2-5 days. GFP or Luciferase may be quantified 48 hours after infection.

ViroVision™ Complete Media Guide

Cell Line	Media
Rev-A3R5	ViroVision™ Growth Media A: RPMI-1640 containing 10-15% FBS, 1% L-Glut, 1% Pen/Strep, and 1 mg/mL Geneticin (G418)
Rev-A3	ViroVision™ Growth Media B: RPMI-1640 containing 10-15% FBS, 1% L-Glut, 1% Pen/Strep
Rev-CEM	ViroVision™ Growth Media C: RPMI, 10% FBS